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Clinical, cognitive and neuroanatomical associations of serum NMDAR autoantibodies in people at clinical high risk for psychosis

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Declaration

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Contribution

TP & PM conceived the study. TP performed analyses and wrote the manuscript.

MK coordinated data collection across EU-GEI sites, analysed neuroimaging data and contributed to the manuscript.

CI, DAM, EC, LJ, RY performed analyses and edited the manuscript.

AV and SI supervised analyses, assisted with interpretation and edited the manuscript.

LdH, SR, GS, AR-R, M-OK, BN, BG, NVB, BR, JvO, RAB, MvdG, LV, MH, SH led participant recruitment at their sites and edited the manuscript.

JS, AD and PM oversaw data collection, assisted with study design and data interpretation and edited the manuscript.

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Abstract

Serum neuronal autoantibodies, such as those to the NMDA receptor (NMDAR), are detectable in a subgroup of patients with psychotic disorders. It is not known if they are present before the onset of psychosis or whether they are associated with particular clinical features or outcomes. In a case-control study, sera from 254 subjects at clinical high risk (CHR) for psychosis and 116 healthy volunteers were tested for antibodies against multiple neuronal antigens implicated in CNS autoimmune disorders, using fixed and live cell-based assays (CBAs). Within the CHR group, the relationship between NMDAR antibodies and symptoms, cognitive function and clinical outcomes over 24 month follow-up was examined. CHR subjects were not more frequently seropositive for neuronal autoantibodies than controls (8.3% vs 5.2%; OR = 1.50; 95% CI: 0.58 – 3.90; $p = 0.40$). The NMDAR was the most common target antigen and NMDAR IgGs were more sensitively detected with live versus fixed CBAs ($p < 0.001$). Preliminary phenotypic analyses revealed that within the CHR sample, the NMDAR antibody seropositive subjects had higher levels of current depression, performed worse on the Rey Auditory Verbal Learning Task ($p < 0.05$), and had a markedly lower IQ ($p < 0.01$). NMDAR IgGs were not more frequent in subjects who later became psychotic than those who did not. NMDAR antibody serostatus and titre was associated with poorer levels of functioning at follow-up ($p < 0.05$) and the presence of a neuronal autoantibody was associated with larger amygdala volumes ($p < 0.05$). Together, these findings demonstrate that NMDAR autoantibodies are detectable in a subgroup of CHR subjects at equal rates to controls. In the CHR group, they are associated with affective psychopathology, impairments in verbal memory, and low IQ: these findings are qualitatively and individually similar to core features of autoimmune encephalitis and/or animal models of NMDAR antibody-mediated CNS disease. Overall the current work supports further evaluation of NMDAR autoantibodies as a possible prognostic biomarker and aetiological factor in a subset of people already meeting CHR criteria.

Introduction

Neuronal surface autoantibodies (NSAbs) have been identified in blood samples from a minority of patients with psychotic disorders and several other unrelated neurological and psychiatric conditions (1-3). These autoantibodies target cell-surface proteins, such as the NMDA receptor (NMDAR), GABA_A or GABA_B receptor, AMPA receptor, LGI1 or CASPR2 or intracellular proteins, such as GAD. When present in CSF, these IgG-subclass cell-surface targeting antibodies are considered pathogenic and associate with severe autoimmune encephalopathy syndromes which can present with varied psychiatric features, particularly in the case of NMDAR antibody encephalitis, plus impaired cognitive function and pathological changes centred around limbic brain regions (4-8). Estimates of the prevalence of NSAbs in patients with psychosis vary depending on the age, condition studied and the assay used. Live cell-based assays (CBAs) consistently detect higher rates than fixed CBAs, and fixed CBA positivity rates vary widely (1, 9-11). Therefore, the clinical relevance of these serum-only antibodies remains unclear.

At present, there are no data on whether NSAbs are present prior to the onset of psychosis and, if so, whether they predict clinical outcomes. People at clinical high risk (CHR) for psychosis experience subthreshold psychotic symptoms, display memory impairments (12) and show alterations in the structure of limbic brain areas (12-14), findings that show a partial resemblance to those seen in NMDAR antibody mediated encephalitis (4, 15-17).

Outside of the autoimmune encephalitis context there is increasing evidence that NMDAR antibodies may be associated with cognitive impairment (e.g. in cancer (18, 19), or following viral encephalitis(20)) but this association has not been examined in the context of psychosis-spectrum disorders. In patients who have developed psychosis, most studies have not found clinically significant differences in symptom profile between seropositive and seronegative patients (3, 9, 10, 21), raising potential doubts over their pathogenic relevance. However, *in vitro*, serum NMDAR antibodies from patients with psychosis disrupt receptor dynamics (22, 23) and function (9, 24), suggesting that the antibodies have pathogenic potential at a molecular level.

It is known that NR1-specific intrathecal B cells – which have not been reported in psychiatric disorders – are a hallmark of NMDAR-antibody encephalitis (25) and that circulating B cells can produce these antibodies (26). The clinical significance of serum autoantibodies may depend on the presence of additional factors, such as blood-brain barrier (BBB) disruption (9, 27), which could allow them access to the CNS. There is widespread evidence for BBB disruption in psychotic disorders (28), although human studies are limited by the requirement for CSF samples or the use of proxy clinical markers such as a history of brain injury. One such marker is the astrocytic calcium-binding protein S100B which is elevated in schizophrenia (28-31), and can be measured in serum samples alongside autoantibodies.

If autoantibodies to the NMDAR play an aetiological role in some cases of psychosis, one would expect that they would be present before the full clinical expression of the disorder. 10-35% of patients with CHR may subsequently develop psychosis, with the proportion varying between samples (32, 33), and there is an unmet clinical need for biomarkers that can predict which individuals will later become psychotic (34). Therefore, we sought to address this by investigating various NSAbs in people who are at CHR for psychosis and examining the relationship between NMDAR antibodies and clinical outcomes, cognitive function and psychopathology in CHR subjects.

Materials and methods

Samples

Sera were taken from 254 subjects from the EU-GEI high-risk study who met the PACE criteria for the CHR state (35), and from 116 healthy controls (HC). CHR subjects were recruited from 11 centres (London, Amsterdam, The Hague, Copenhagen, Vienna, Basel, Cologne, Melbourne, Paris, Barcelona, and São Paulo) and were reassessed at a time point as close to 12 and 24 months as possible. Exclusion criteria were presence of a current or past psychotic disorder, symptoms relevant for inclusion being explained by a medical disorder or drugs or alcohol dependency or IQ < 60.

116 HC subjects, frequency matched by age and sex, were recruited from the EU-GEI high-risk study (n=55), and the South East London Community Health (SELCOH) study (36) (n = 61). HCs from the EU-GEI study were excluded if they met criteria for the CHR state, while HCs from SELCOH were excluded if they screened positive on the Psychosis Screening Questionnaire (37). Ethical approval for the study was obtained from the local research ethics committees at each of the sites. All participants provided written informed consent.

All serum samples were stored at -80 degrees Celsius until assayed and tested blind to case-control status.

Phenotype data

All subjects completed a sociodemographic schedule. Baseline psychopathology was assessed using the Comprehensive Assessment of At-Risk Mental States (CAARMS) (38), Brief Psychiatric Rating Scale (BPRS) (39), Scale for the Assessment of Negative Symptoms (SANS) (40), Young Mania Rating Scale, (YMRS) (41), and the Montgomery-Åsberg Depression Rating Scale (MADRS) (42). The Rey Auditory Verbal Learning Task (43) was used to assess verbal memory function and IQ was estimated using the shortened Wechsler Adult Intelligence Scale III (44). Overall social functioning was assessed using the Global Assessment of Functioning (GAF) scale. Participants were invited for face-to-face follow-up meetings at 1 year and 2 years after baseline. Where face-to-face meetings were not

possible, participants were followed up for 2 years using available clinical records, and this follow-up was extended when additional clinical data was available.

Immunoassays

Sera were tested for antibodies of the IgG, IgA and IgM subtypes to 32 different substrates (see supplementary appendix 1 for a full list), by indirect immunofluorescence on cell-based assays using fixed recombinant HEK293 cells and on frozen sections of rodent and monkey brain tissue (hippocampus and cerebellum) at Euroimmun AG, Lübeck, Germany, as described previously (45)).

Live CBA autoantibody testing for the detection of IgG antibodies to NMDAR was performed using HEK293 cells (European Collection of Authenticated Cell Cultures (ECACC), Public Health England) at the Nuffield Department of Clinical Neurosciences, University of Oxford as described previously (46) with some modifications. Specifically, intensity of fluorescence of each sample was rated on a semi-quantitative visual scoring system of 0-4. Scores of 0 and 1 were rated as negative, 1.5 as low positive and 2-4 as positive. All positive samples were confirmed at dilutions of 1:20, 1:100 and 1:500 using an IgG-Fc specific secondary antibody (goat antihuman Fc IgG; Thermo Scientific 31 125), then tested for non-specific binding using HEK293 cells that had been transfected with another antigen (dopamine D2 receptor). The titre of the antibody was given as the dilution of serum giving a score of 1. IgG positive samples were tested in the Oxford laboratory for binding to live, cultured rat hippocampal neurons as described previously (47). All cell lines are regularly tested for *Mycoplasma* contamination using immunofluorescence.

Levels of S100B, a putative marker of blood-brain barrier disruption, were assessed using a chemiluminescence assay (Diasorin). High-sensitivity C-reactive protein (hsCRP) was measured as a general marker of systemic inflammation, using high sensitivity enzyme immunoassays as previously described (48).

Neuroimaging analysis

MR Images were acquired using the ADNI-2 T1 weighted volumetric sequence (<http://adni.loni.usc.edu/methods/documents/mri-protocols/>) from 160 CHR subjects on 3T scanners. The ADNI-2 sequence has been specifically developed for multicentre studies and has been optimised to give similar contrast across MRI scanners. Nevertheless, the scanner was modelled as a factor in the statistical analysis to control for this. T1 weighted volumes were and preprocessed using Freesurfer version 6.0.0, using a standard pipeline for quantitation of subcortical volumes (described in full on the FreeSurferWiki page (<https://surfer.nmr.mgh.harvard.edu/fswiki>)). First, intensity correction and skull stripping were performed, followed by segmentation of grey and white matter along with segmentation of subcortical structures in which subject scans are warped to a training set atlas and individual voxels labelled using a Gaussian classifier analysis, yielding maps of deep grey matter, white matter, and CSF structures.

Analyses were performed on Freesurfer's subcortical segmentation output volumes and estimates of total intracranial volumes. Analysis was *a priori* restricted to the hippocampus and the amygdala (both bilaterally), as these are the areas that are predominantly affected in autoimmune encephalopathies associated with NSAbs, and these areas have been independently linked to the later onset of psychosis in neuroimaging studies of clinical high-risk populations (49, 50).

Statistical analysis

Statistical analyses were performed using SPSS 23. Demographic variables were compared between cases and controls and between seropositive and seronegative CHR subjects with independent samples t-tests and Mann-Whitney U tests for continuous variables, and chi square and Fisher's exact tests for categorical data. Based on previous work on neuronal autoantibody prevalence in first episode psychosis populations(10), sample sizes were selected to have statistical power to detect differences.

For symptom and cognitive scales analyses of variance was performed within the General Linear Model with NMDAR antibody status (positive or negative), gender and ethnicity as

factors and with age as covariate, after homogeneity of variance was confirmed. Given the polymorphic psychiatric presentation in NMDAR antibody encephalitis (7, 51) we aimed to look at associations with a broad range of psychopathology rather than restricting analyses to any one symptom domain or dimension. In addition to total scores for the CAARMS (whole instrument score), YMRS, MADRS and SANS, a positive symptoms factor was computed based on the BPRS components analysis of Dingemans et al. (52).

For dichotomous outcome measures (transition, nonremission from CHR state), logistic regression was performed with NMDAR antibody status (positive or negative), age, gender and ethnicity as covariates.

To compare subcortical 'limbic' volumes between seropositive and seronegative CHR subjects multivariate analysis of variance with NSAb status (positive or negative), MRI site, gender and ethnicity as factors, and total intracranial volume (TICV) and age as covariates, was performed in the General Linear Model. All main effects were included in our models and the analysis of interactions was *a priori* limited to the interaction between NSAb status and S100B status.

Significance threshold was set to $p < 0.05$; mean \pm sd presented. Due to the exploratory nature of the analyses, significance values are given for two-tailed tests where applicable and uncorrected for multiple comparisons.

Results

Participants

Demographic and clinical information is presented in Table 1. Cases and controls differed in ethnicity, with fewer black participants in the CHR group (8.7%) than in the healthy control group (20.7%). The groups did not differ on any other demographic variable. There was no difference in mean serum S100B level between CHR subjects and controls ($p = 0.833$); CHR subjects had slightly higher baseline inflammation as indexed by hsCRP ($p = 0.048$). Smoking and antipsychotic usage was more frequent in cases than in controls.

Prevalence of NSAbs detected using commercial, fixed CBA and live CBA

Using commercial, fixed assays, NSAbs were detected in 21 (8.3%) CHR subjects and 6 (5.2%) of matched, healthy controls (OR = 1.50; 95% CI: 0.58 – 3.90; $p = 0.401$). Prevalence did not differ by site ($p = 0.800$). IgG was the most commonly detected antibody isotype (48.1%) followed by IgA (33.3%) and IgM (22.2%). Isotype distribution did not differ between CHRs and HCs (IgG: $p = 1.000$; IgA: $p = 0.283$; IgM: $p = 0.670$). See Figure 1c for distribution of all NSAb targets. NMDAR was the most commonly detected antigen in the CHR group ($n = 11$ [4.3%]; comprising IgG = 1 [0.4%], IgA = 6 [2.4%] and IgM = 4 [1.6%]) and in the HC group ($n = 2$ [1.7%]; comprising IgG = 1 [0.9%], IgA = 0 and IgM = 1 [0.9%]) (Figure 1d). There was no difference in overall NMDAR antibody seroprevalence between the CHR and HC groups ($p = 0.36$). Although NMDAR antibody isotype distribution was not different between the two groups (IgG: $p = 0.529$; IgA: $p = 0.183$; IgM: $p = 1.000$), IgA NMDAR antibody seropositivity was seen in 6 CHR subjects but not in any HC.

On fixed CBAs, titres to specific antigens ranged from 1:10 to 1:320 in CHR subjects and from 1:32 to 1:320 in HCs (Figure 1e), but none of these bound brain tissue slices on immunohistochemistry conducted in the same laboratory. However, 11 of 12 (91.7%) NSAb IgG seropositive samples bound cultured rat hippocampal neurons, with three samples showing strong binding. Serum from seven subjects (5 CHR subjects, 2 HCs) had IgG, IgA or IgM antibodies which bound cerebellum or hippocampus sections (titre 1:32 to 1:320) without antigen-specific antibodies detected. No subjects were seropositive for more than one antigen on fixed CBA.

Since studies that have used live CBA to detect NMDAR IgG in psychotic patients and controls have tended to yield greater seropositivity rates (23) and find case-control differences (10, 11, 22), indicating greater analytical sensitivity, we retested all samples for NMDAR IgG using a live CBA (Figure 2a). Live CBA detected a significantly higher prevalence of NMDAR IgG antibodies in the total cohort than did fixed CBA (19 (5.1%) IgG seropositive subjects on live CBA vs 2 (0.5%) on fixed CBA: $p = 0.0001$), but were found similarly in 5.1% of CHR cases and 5.2% of HCs ($p = 1.000$). Prevalence did not differ by site ($p = 0.145$). Titres ranged from 1:30 to 1:750 in CHR subjects and from 1:40 to 1:500 in HCs (Figure 2b). The

two subjects who were seropositive for NMDAR IgG on fixed CBA were also seropositive on live CBA.

17 of 19 samples which were NMDAR IgG seropositive on live assays were tested for binding to cultured rat hippocampal neurons: 10 of 17 (58.8%) samples bound neurons, with no samples showing strong binding. All four samples with a CBA titre of 1:250 or over bound neurons; of the 13 samples with lower titres, six (46.2%) bound neurons. IgG titre on CBA and hippocampal binding intensity score were positively correlated ($r = 0.491$; $p = 0.048$).

There were no age, sex, ethnicity or smoking status differences between seropositive and seronegative subjects (table 2). S100B and hsCRP levels did not differ between seropositive and seronegative CHR subjects, for any NSAb or for NMDAR antibodies measured using either assay (table 2) .

There were no associations between antibody serostatus and self-reported medical history including neurological disorder, autoimmune disorder, head trauma, cancer, history of surgical procedures, CNS infection or serious infection (supplemental table 1). Notably the only subject that had had a CNS infection (meningitis) was seropositive for NMDAR IgG on live assay.

Clinical outcomes of CHR subjects

Of the 254 CHR subjects from whom serum had been collected and tested for NSAbs, 49 (19.3%) developed a psychotic illness in the follow-up period. The subjects who transitioned to psychosis did not differ from those who did not transition in age or gender (Supplemental Table 2).

Of the subjects who transitioned to psychosis, 1 (2.0%) was seropositive for NMDAR antibodies on fixed CBA and 3 (6.1%) were seropositive for NMDAR IgG on live assay at baseline; of 205 CHR subjects who did not transition to psychosis, 10 (4.9%) were seropositive on fixed assay and 10 (4.9%) were seropositive on live assay (Supplemental Table 3). In logistic regression analyses, NMDAR antibody serostatus at baseline did not predict later transition to psychosis or nonremission from the CHR state, regardless of the assay used (fixed or live CBA) for antibody detection (Supplemental Tables 4-7).

Scores of functioning were assessed in CHR subjects at the assessment timepoints closest to two years after baseline. Mean change in GAF scores was negative (indicating worsening function) for seropositive subjects (live assay), and positive (indicating improvement) in seronegative subjects; in multiple regression analyses NMDAR antibody seropositivity was significantly associated with change in GAF disability scores (Supplemental Table 8). There was a significant negative correlation between NMDAR IgG titre (live assay) and change in GAF disability scores (Spearman's $\rho = -0.542$; $p = 0.024$), with higher titre subjects showing a more negative change in GAF scores, indicating deteriorating function.

Cognitive associations of NMDAR antibody seropositivity in CHR subjects

Total RAVLT immediate recall score was lower in CHR subjects seropositive for NMDAR antibodies on fixed CBA compared to seronegative CHR subjects ($p = 0.030$). These differences became more substantial after controlling for total CAARMS score ($p = 0.007$), suggesting that the relationship with auditory verbal memory was not simply secondary to an effect of the severity of psychopathology (Table 3).

In order to explore whether the association with cognitive impairment was specific to verbal memory function, we then compared IQ (estimated using the WAIS-III) in seropositive and seronegative subjects. Mean WAIS-III IQ in NMDAR antibody seropositive CHR subjects was almost 15 points lower than that in seronegative CHR subjects ($p = 0.010$). This group difference also remained significant after controlling for total CAARMS score ($p = 0.008$) (Table 3). Both effects appeared to be driven by IgA/M antibodies: after restricting the analysis to IgA/M seropositive subjects only, both AVLT immediate recall ($p=0.011$) and WAIS-III estimated IQ ($p= 0.018$) remained significantly impaired in seropositive subjects.

In fact, within the total CHR sample ($n=254$), only 9 subjects had an IQ below 70 (the threshold for a diagnosis of intellectual disability) but 3 of these CHR subjects (33%) were seropositive for NMDAR antibodies. Subjects who were NMDAR IgG seropositive on live CBA did not differ from seronegative subjects on measures of cognition including AVLT immediate recall ($p = 0.337$) and IQ ($p = 0.796$).

Symptomatic associations of NMDAR antibody seropositivity in CHR subjects

Nine of 13 (69.2%) subjects who were seropositive for NMDAR antibodies on live assay met DSM-IV criteria for current depressive episode, compared to 31.2% of seronegative subjects ($p = 0.012$). Rates of other DSM-IV diagnoses did not differ according to serostatus. There was no difference in the rates of specific DSM-IV diagnoses between NMDAR antibody fixed CBA seropositive and seronegative subjects (supplemental table 9).

While symptom severity scores were not significantly different between seropositive and seronegative subjects for most symptom domains when looking at antibodies detected using either assay, MADRS scores were borderline significantly higher in CHR subjects seropositive on live CBA ($p = 0.051$), indicating greater depression symptom severity. Across the whole CHR cohort, MADRS scores were weakly correlated with NMDAR IgG (live CBA) titre (Spearman's $\rho = 0.167$; $p = 0.009$).

There was no association between NMDAR antibody serostatus and CHR subtype (vulnerability group, APS or BLIPS) (fixed CBA: $p = 0.316$; live CBA: $p = 0.865$), although the numbers of subjects in the vulnerability ($n = 41$) and BLIPS ($n = 16$) subgroups were small.

Association between blood-brain barrier disruption and symptoms

In order to assess the possible role of blood-brain barrier disruption as a moderating variable for the associations of NMDAR serostatus with symptom measures, we performed a preliminary (due to small n in the seropositive groups) analysis of the association between serum S100B levels and symptom scores in NMDAR antibody seropositive and seronegative groups. In subjects for whom NMDAR IgG were detected using live CBA, positive associations between S100B and symptom severity at baseline were observed for total BPRS score and for negative symptoms. In the seronegative group, modest negative associations between S100B and symptom severity were observed for multiple symptom scales (supplementary table 10).

Longitudinal NSAb serostatus

179 follow-up serum samples were available on 120 CHR subjects (modal interval between baseline and repeat blood sample: 12 months). Baseline serostatus was predictive of follow-up serostatus i.e. subjects who were seropositive at baseline were more likely than subjects who were seronegative at baseline to be seropositive at follow-up. In total, of 109 CHR subjects who were seronegative at baseline and who had blood taken at subsequent visits, 9 (8.3%) were seropositive at subsequent visits (i.e. had new neuronal antibodies detectable). Of 11 subjects who were seropositive at baseline, 4 (36.4%) were still seropositive on at least one subsequent visit ($p = 0.018$). In 3 of these 4 subjects (75%), NSAbs detectable at follow-up were to the same antigen and of the same isotype.

None of the four subjects with persistent antibodies (i.e. NSAbs detectable at baseline and at a subsequent timepoint) became psychotic during follow-up. Only one of 9 subjects who developed *de novo* NSAbs at follow-up became psychotic: this subject developed NMDAR IgM antibodies (1:10) a year after initial assessment, having been seronegative at baseline; he went on to transition to psychosis a year later, but no blood samples were available at the time he became psychotic or afterwards.

Of 5 subjects who had NMDAR antibodies detectable at baseline and on whom follow-up samples were available, two (40%) had NSAbs at one or more follow-up point. One case with NMDAR IgA antibodies (1:32) at baseline had IgG antibodies at visit 2 (12 months) which bound hippocampus and cerebellum (1:100) but no NMDAR-specific antibodies detectable, and at visit 4 (24 months) had no antibodies at all detectable. One case with NMDAR IgM antibodies at baseline (1:100) had NMDAR IgM antibodies (1:100) detectable at visit 3. Neither of these cases developed psychosis.

Of 11 CHR subjects on whom serum was taken at the time of transition to psychosis, none were seropositive.

Limbic volumes in NSAb seropositive CHR subjects

Because only a small number of the CHR subjects who were NMDAR antibody seropositive had also had an MRI scan (n=5), we investigated the relationship between autoantibodies and MRI measures in subjects who were seropositive for any NSAb (n=11). CHR subjects who were seropositive for NSAbs had larger amygdala volumes ($p = 0.048$) than seronegative CHR subjects (n=148). There were no group differences in hippocampal volume (supplementary table 11).

Discussion

Summary of results

This is the first study of CNS autoantibodies in people at CHR for psychosis. We found an overall seroprevalence of 8.3%, indicating that NSAbs are detectable in a minority of these individuals. The NMDAR was the most common target antigen and NMDAR IgGs were more sensitively detected with live versus fixed CBAs. The prevalence was not significantly greater than in HCs, mirroring findings from recent studies that have found similar case-control seroprevalence rates in patients with psychosis and other psychiatric and non-psychiatric disorders (1, 21, 53). NMDAR antibody seropositivity was not associated with the development of a psychotic disorder, which was our main outcome of interest, but was associated with a deterioration in disability-associated functional status. Within the CHR sample, NMDAR antibody seropositivity was associated with impaired verbal memory performance, lower IQ and with depressive symptoms. Exploratory MRI analysis indicated that NSAb seropositivity was associated with increased amygdala volume.

Relevance of NSAbs in CHR subjects

The combination of impaired cognitive function, severe affective psychopathology (46, 51, 54) and increased volume of limbic brain regions (55-58) bears some resemblance to (but is less severe than) the pattern of findings seen in the acute stages of many autoimmune encephalopathies. This suggests that NMDAR antibodies, and possibly other NSAbs, could play an analogous, but not necessarily identical, role in the pathophysiology of some patients who present with psychiatric symptoms (59). This would be consistent with recent

evidence that NMDAR antibodies isolated from psychiatric patients have pathogenicity *in vitro* and *in vivo* (9, 22, 24).

The reductions in verbal memory performance in NMDAR antibody positive subjects are reminiscent of those seen in NMDAR encephalitis, and were of a comparable magnitude (19). It is notable however that seropositive CHR subjects did not show other features of autoimmune encephalopathies, such as seizures or frank movement disorders. Neurological soft signs were not assessed in the present study, but subtle movement signs have been observed in a proportion of CHR subjects (60-62) and have been associated with NMDAR antibody seropositivity in patients with psychosis (9).

Furthermore, NMDAR-antibody encephalitis is classically associated with a polymorphic affective psychosis with catatonic features (7), which is broader than the phenotypic associations of seropositivity that we observed. However a rodent model involving the intraventricular administration of NMDAR antibodies generated a cognitively impaired, anxious and depressive phenotype (63), which is consistent with our findings of an association between NMDAR antibodies and MADRS scores and current depressive episodes in CHR subjects. It has been suggested that the multifaceted symptomatology of NMDAR encephalitis represents more than the effects of the NMDAR antibody at the synapse, and that additional immune and non-immune factors help shape clinical presentation. Most passive immunisation animal models do not recapitulate a full encephalopathic phenotype, instead showing convergence around a more restricted set of symptoms which could be conceived as a 'ketamine-like' synaptopathy (64), without features of frank CNS inflammation. If NMDAR antibodies do have a causal role in some psychiatric disorders, it is therefore possible that the effects represent more of a synaptopathic than a frank encephalitic process.

Additionally, the possibility that NMDAR antibodies could play a causal role in some CHR cases does not imply that the overall underlying immunology in these cases would be the same as in NMDAR encephalitis, a disorder which is characterised by the presence of NR1-specific intrathecal B cells. Indeed the importance of BBB disruption in determining the relevance of peripheral NMDAR antibodies in many human and animal studies of NSAb-

mediated psychiatric disorders (9, 27, 64)) suggests that intrathecal synthesis may be minimal or absent in seropositive subjects with isolated psychiatric presentations.

Given the associations we report between NSAbs and both amygdala enlargement and affective symptomatology, it is interesting to note that whereas amygdala volumes are reduced in schizophrenia and first episode schizophrenic psychosis, they are increased in the first episode of affective psychoses and other nonschizophrenic psychoses (including brief psychotic disorder) (65). The increase in amygdala volumes in our NSAb seropositive subjects could represent acute inflammation, as is the case in acute limbic encephalitis. Importantly, in the post-acute phase of autoimmune encephalitis there is evidence of volume loss in these same regions, which is in turn linked to the severity of persisting deficits (5, 15, 17). If the neuroanatomical associations of NSAb seropositivity truly represent a similar mechanism to that of autoimmune encephalitis, one testable hypothesis is that these brain regions will show accelerated atrophy or volume loss at follow-up scans in seropositive CHR subjects.

Prüss, Finke and colleagues have highlighted associations between IgA NMDAR antibodies and cognitive impairments in multiple clinical contexts (2, 18, 66). In the present study, 6 CHR subjects had IgA NMDAR antibodies, compared with no HCs. It is thus possible that IgA NMDAR antibodies drive the cognitive impairment observed in our seropositive subjects. IgM, and many IgAs, are not frequently reported in the CSF, although this may potentially reflect a bias towards the measurement of IgG only. While IgA can exist as monomers in blood, IgMs are multimerised antibodies, potentially making their crossing of the blood brain barrier less likely than IgGs.

We observed that in CHR individuals NMDAR antibody serostatus fluctuates over time. Pan and colleagues recently reported similar findings in animals and human subjects (following stroke): a majority of individuals remained seronegative, and a minority either remained seropositive or acquired *de novo* antibodies at 1-3 year follow-up (67). What causes these fluctuations, and whether they have any direct clinical consequences, has not been established (although there is some emerging evidence for chronic stress as a potential inducer of NMDAR antibodies).

Comparison of cell-based assays

In the largest direct assay comparison reported to date, we found that live CBA detects a nine times higher prevalence of IgG NMDAR antibodies in serum than fixed CBA, confirming a previous report in a first episode psychosis cohort (23). In this earlier report, even in the absence of seropositivity on fixed CBA, these IgG both targeted and were capable of altering the surface dynamics of the NMDAR suggesting pathogenic potential.

In total 72.4% of IgG seropositive samples bound hippocampal neurons; this was more true of IgG seropositive samples on fixed CBA (91.7%) than on live CBA (58.8%). This is consistent with live CBA potentially detecting antibodies with higher sensitivity than the fixed CBA. The fact that in our laboratory not all live CBA-positive samples bound rat hippocampal neurons (see also (21)) does not undermine the potential disease-relevance of NMDAR IgG seropositivity, particularly as these antibodies have pathogenic potential at the level of the NMDAR (22, 23). Immunocytochemistry with cultured neurons, as with all assays, has a detection threshold, and this should be understood in the context that CBAs overexpress the antigen at the surface but in immunocytochemistry expression levels are broadly physiological. Therefore it might be expected that some lower titre samples that are positive on live assay will not demonstrate binding to neurons. The observed correlation between NMDAR IgG live CBA titre and hippocampal binding intensity is suggestive that live CBA is measuring genuine IgG binding, and indeed the live CBA threshold above which NMDAR IgG positive samples will reliably bind to neurons appears to be 1:250.

The role of the blood-brain barrier

Ehrenreich and colleagues have stressed that for peripheral NSAbs to be clinically relevant there may also need to be disruption of the BBB (64). This has been demonstrated in mice with an ApoE^{-/-} haplotype (68) and in humans with a 'history of neurotrauma' or obstetric complications as medical proxies (9). A parsimonious interpretation is that when the integrity of the BBB is compromised, NSAbs have greater access to the CNS and are more likely to disrupt brain function (28). However, it is also possible that serum NSAbs are

peripheral biomarkers of a central dysfunction that is accompanied by BBB disruption, such as neuroinflammation. Consistent with the role of the BBB as proposed by Ehrenreich and colleagues, in a supplemental analysis we observed positive associations between S100B levels (a widely used proxy for BBB disruption (28, 31)) and symptom scores in subjects who were seropositive for NMDAR antibodies on live CBA; these associations were in the opposite direction in subjects without detectable serum antibodies. Given the small numbers of seropositive subjects however this finding mandates replication in a larger sample, potentially with an additional marker of BBB disruption. The 'gold standard' of BBB quantification in humans is the CSF-serum albumin quotient, which has consistently been found to be abnormal in psychosis (28). However this requires a lumbar puncture, which is not usually part of the clinical assessment of patients with psychosis, let alone individuals at CHR (69). A less invasive method for BBB quantification in these individuals is the use of dynamic contrast-enhanced MRI (70).

Antibodies as biomarkers in the CHR state

Separately, or in addition to, the question of the pathogenicity of NMDAR antibodies in the CHR state, the present study indicates a potential role for these antibodies as biomarkers of poor cognition, depressive symptomatology and/or outcome in the CHR state.

In CHR subjects, severe psychopathology and cognitive impairments, and alterations in limbic brain volumes at presentation are associated with an increased risk of later transition to psychosis (12-14, 71, 72). Impaired verbal memory performance in CHR individuals is one of the strongest cognitive predictors of subsequent transition to psychosis (73, 74). Low IQ has also been identified as a predictor of conversion to psychosis in CHR subjects (75), and is an established risk factor for psychosis in the general population (76). Our study suggests, complementing similar findings in other patient groups (2, 18, 20, 66), that NMDAR antibodies may have a role as a biomarker of cognitive impairment in CHR subjects. Since we observed that antibody serostatus fluctuates over time, an important question is whether cognitive status fluctuates in association with serostatus. This latter might be expected if NMDAR antibodies represent state (rather than trait) biomarkers for poor cognition: indeed most passive transfer models suggest that the antibody effects are

reversible. However a different relationship might be expected if serostatus is in fact a marker of poor cognitive performance due to the impact of an as-yet undetermined confounder.

In our study, NMDAR antibodies did not predict transition to psychosis, as defined by operationalised criteria, or nonremission from the CHR state, despite all but one NMDAR IgG seropositive subjects still meeting CHR criteria at follow-up. NMDAR antibody seropositivity was however associated with a deterioration in disability-associated functioning, short of transition, and indeed greater antibody titre was associated with greater deterioration in function, overall suggesting that NMDAR antibody serostatus should be further evaluated as a predictive marker of functional outcome.

Limitations

We measured NSAbs in serum. Assessing NSAbs in cerebrospinal fluid (CSF) may provide a better way of investigating their role in some autoimmune CNS disorders, but lumbar puncture is more invasive and is not usually carried out in CHR subjects. However, while CSF NSAbs are important for the diagnosis of autoimmune encephalitis, this does not imply that serum NSAbs are irrelevant in psychiatric conditions such as the CHR state. As Castillo-Gomez and colleagues have demonstrated, in conditions in which antibody production is putatively lower than in autoimmune encephalitis the brain may act as ‘immunoprecipitator’ of CSF NSAbs, and NSAbs may not be detectable in CSF, despite having functional consequences in the brain (68). Although we studied a large sample of CHR subjects, because only a minority were antibody-positive, the numbers of subjects in some of the comparisons was small and hence we did not perform corrections for multiple comparisons. We also cannot exclude the possibility that we failed to detect some differences because of limited statistical power. This issue may be addressed by pooling data from different CHR cohorts, as in the Harmony collaboration (https://www.pronia.eu/news-pronia/news-detail/tx_news/initiation-of-the-multi-center-collaboration-harmony-in-october-copy-1/).

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Figure legends

Figure 1. NSAb seroprevalence and associations of seropositivity in CHR and HCs – antibodies detected using fixed assay. **a)** Examples of seropositive immunoassays. Top left: fixed CBA seropositive for NMDAR antibody IgG (CHR subject); human IgG is labelled green using a fluorescent secondary antibody. Top right: immunohistochemistry showing IgG binding to fixed and permeabilised rat hippocampus (CHR subject seronegative for other specific antigenic targets); human IgG is labelled green using a fluorescent secondary antibody. Bottom left: IgG from CHR subject showing strong binding to cultured hippocampal neurons (score 3 out of 4); human IgG is labelled green using a fluorescent secondary antibody (AlexaFluor 488 mouse anti-human IgG1; Invitrogen) (left and right) and hippocampal neurons are labelled red using mouse MAP2 antibodies (Monoclonal-Anti-MAP2, clone HM-2, Sigma Aldrich) (right). Scalebar = 25um. **b)** Confocal microscope image of IgG from CHR subject showing strong binding to cultured hippocampal neurons. Cell nuclei are DAPI-labelled; human IgG is labelled green using a fluorescent secondary antibody (AlexaFluor 488 mouse anti-human IgG1; Invitrogen) and hippocampal neurons are labelled red using mouse MAP2 antibodies (Monoclonal-Anti-MAP2, clone HM-2, Sigma Aldrich). Scalebar = 25um. **c)** Distribution of antigen target by group (note percentages are of the totality of samples tested). **d)** Distribution of antibody isotype by group. **e)** Distribution of antibody titre by group and assay type; IgG = green, IgA = red, IgM = blue.

Figure 2: NMDAR IgG seroprevalence and associations of seropositivity – antibodies detected using live assay. **a)** Representative example of NMDAR IgG positive live CBA. Human IgG is labelled red using a fluorescent secondary antibody (AlexaFluor 568 goat anti-mouse IgG (H+L); Invitrogen) and shown binding to cells that co-express eGFP (green). Scalebar = 25um. **b)** Distribution of NMDAR IgG titre by group and assay type (fixed vs live assay).

Table 1: Demographic/basic clinical information and NSAb serostatus by group

*data available for 365 of 370 subjects

**Adjusted for BMI, gender, age, ethnicity and smoking status

Abbreviations: CHR: clinical high risk, HC: healthy control, CBA: cell-based assay, S100B: S100 calcium-binding protein B, CRP: C-reactive protein, NMDAR: N-methyl-d-aspartate receptor.

Table 2: Demographic/basic clinical information by NSAb serostatus

*data available for 365 of 370 subjects

**Statistic adjusted for BMI, gender, age, ethnicity and smoking status

Abbreviations: CBA: cell-based assay, S100B: S100 calcium-binding protein B, CRP: C-reactive protein, NMDAR: N-methyl-d-aspartate receptor, BMI: body mass index.

Table 3: Cognitive function and psychopathology scores by NMDAR antibody serostatus (descriptive means shown with F and p values from ANOVA). Numbers in brackets under variable names are the number of seropositive subjects and seronegative subjects for whom data were available for that variable. BPRS total score excluded from analysis due to heteroscedasticity.

| | Total CHR cohort (n=254) | Total HC cohort (n=116) | p |
|------------------------------------|---|---|------------------|
| Age | 22.70 ± 5.00 (range 14 to 45) | 23.54 ± 3.38 (range 17 to 34) | 0.057 |
| Sex [M: n (%)] | 136 (53.5) | 60 (51.7) | 0.822 |
| Ethnicity [n (%)] | White 178 (70.4), Black 22 (8.7), Other 53 (20.9) | White 76 (65.5), Black 24 (20.7), Other 16 (13.8) | 0.003 |
| Smoker* [n (%)] | 133 (53.4) | 31 (26.7) | <0.001 |
| BMI | 24.31 ± 5.30 | 23.70 ± 4.44 | 0.288 |
| Current antipsychotic use [n (%)] | 24 (9.4) | 0 (0) | <0.001 |
| S100B (ug/L)** | 0.040 ± 0.049 | 0.050 ± 0.071 | 0.833 |
| CRP (mg/L)** | 1.23 ± 1.08 | 0.95 ± 0.95 | 0.048 |
| Any antibody (fixed CBA) [n (%)] | 21 (8.3) | 6 (5.2) | 0.390 |
| IgG [n (%)] | 9 (3.5) | 4 (3.4) | 1.000 |
| IgA [n (%)] | 8 (3.1) | 1 (0.9) | 0.283 |
| IgM [n (%)] | 5 (2.0) | 1 (0.9) | 0.670 |
| NMDAR antibody (fixed CBA) [n (%)] | 11 (4.3) | 2 (1.7) | 0.360 |

| | | | |
|-------------------------------------|----------|---------|-------|
| <i>NMDAR IgG [n (%)]</i> | 1 (0.4) | 1 (0.9) | 0.529 |
| <i>NMDAR IgA [n (%)]</i> | 6 (2.4) | 0 (0.0) | 0.183 |
| <i>NMDAR IgM [n (%)]</i> | 4 (1.6) | 1 (0.9) | 1.000 |
| <i>NMDAR IgG (live CBA) [n (%)]</i> | 13 (5.1) | 6 (5.2) | 1.000 |

Table 1: Demographic/basic clinical information and NSAb serostatus by group

*data available for 365 of 370 subjects

**Adjusted for BMI, gender, age, ethnicity and smoking status

Abbreviations: CHR: clinical high risk, HC: healthy control, CBA: cell-based assay, S100B: S100 calcium-binding protein B, CRP: C-reactive protein, NMDAR: N-methyl-d-aspartate receptor.

| | <i>Any antibody negative (n=343)</i> | <i>Any antibody positive (fixed CBA) (n=27)</i> | <i>p</i> | <i>NMDAR antibody negative (fixed CBA) (n=357)</i> | <i>NMDAR antibody positive (fixed CBA) (n=13)</i> | <i>p</i> | <i>NMDAR antibody negative (live CBA) (n=351)</i> | <i>NMDAR antibody positive (live CBA) (n=19)</i> | <i>p</i> |
|--|---|---|----------|--|---|----------|---|--|----------|
| <i>Age</i> | 23.01 ± 4.60 | 22.33 ± 4.13 | 0.458 | 22.97 ± 4.57 | 22.69 ± 4.57 | 0.829 | 22.92 ± 4.50 | 23.79 ± 5.73 | 0.418 |
| <i>Sex [M: n (%)]</i> | 178 (51.9) | 18 (66.7) | 0.163 | 187 (52.4) | 9 (69.2) | 0.270 | 188 (53.6) | 11 (57.9) | 0.330 |
| <i>Ethnicity [n (%)]</i> | White 231 (67.5) Black 45 (13.2) Other 66 (19.3) | White 23 (85.2) Black 1 (3.7) Other 3 (11.1) | 0.148 | White 44 (68.5) Black 45 (12.6) Other 67 (18.8) | White 10 (76.9) Black 1 (7.7) Other 2 (15.4) | 0.797 | White 243 (69.4) Black 43 (12.3) Other 64 (18.3) | White 11 (57.9) Black 3 (15.8) Other 5 (26.3) | 0.563 |
| <i>Smoker* [n (%)]</i> | 155 (45.9) | 9 (33.3) | 0.233 | 160 (45.5) | 4 (30.8) | 0.398 | 155 (44.8) | 9 (47.4) | 0.826 |
| <i>BMI</i> | 24.17 ± 4.98 | 23.19 ± 5.55 | 0.370 | 24.07 | 24.94 | 0.591 | 24.12 ± 5.02 | 23.82 ± 5.10 | 0.815 |
| <i>Current antipsychotic use [n (%)]</i> | 21 (6.1) | 3 (11.1) | 0.403 | 22 (6.2) | 2 (15.4) | 0.203 | 22 (6.3) | 2 (10.5) | 0.463 |
| <i>CRP (mg/L)**</i> | 1.14 ± 1.04 | 1.12 ± 1.21 | 0.991 | 1.14 ± 1.04 | 1.28 ± 1.41 | 0.925 | 1.14 ± 1.05 | 1.17 ± 0.94 | 0.506 |

| | | | | | | | | | |
|-----------------------|---------------|---------------|-------|---------------|--------------|-------|---------------|---------------|-------|
| <i>S100B</i> (ug/L)** | 0.043 ± 0.058 | 0.038 ± 0.033 | 0.613 | 0.043 ± 0.057 | 0.04 ± 0.043 | 0.994 | 0.043 ± 0.058 | 0.037 ± 0.031 | 0.783 |
|-----------------------|---------------|---------------|-------|---------------|--------------|-------|---------------|---------------|-------|

Table 2: Demographic/basic clinical information by NSAb serostatus

*data available for 365 of 370 subjects

**Statistic adjusted for BMI, gender, age, ethnicity and smoking status

Abbreviations: CBA: cell-based assay, S100B: S100 calcium-binding protein B, CRP: C-reactive protein, NMDAR: N-methyl-d-aspartate receptor, BMI: body mass index.

| | <i>Antibody positive</i> | <i>Antibody negative</i> | <i>F (serostatus)</i> | <i>p (serostatus)</i> |
|--|------------------------------|------------------------------|-----------------------|-----------------------|
| <i>NMDAR ANTIBODY (IgG, IgA, IgM; FIXED ASSAY)</i> | | | | |
| <i>AVLT IR total (7:160)</i> | 52.00 ± 13.79 | 63.06 ± 11.90 | 4.817 | 0.030 |
| <i>AVLT IR total (CAARMS adjusted) (5:127)</i> | 45.80 ± 10.69 | 62.50 ± 11.90 | 7.651 | 0.007 |
| <i>WAIS estimated total IQ (9:222)</i> | 84.22 ± 20.35 | 98.71 ± 16.90 | 6.687 | 0.010 |
| <i>WAIS estimated total IQ (CAARMS adjusted) (7:182)</i> | 83.29 ± 21.62 | 100.32 ± 16.86 | 7.134 | 0.008 |
| <i>CAARMS total (9:198)</i> | 49.00 ± 16.17 | 49.49 ± 15.00 | 0.001 | 0.981 |
| <i>BPRS positive sx (11:224)</i> | 14.91 ± 5.43 | 13.55 ± 4.54 | 0.731 | 0.393 |
| <i>SANS (9:210)</i> | 20.44 ± 15.30 | 15.53 ± 11.74 | 1.074 | 0.301 |
| <i>YMRS (11:225)</i> | 4.27 ± 5.35 | 4.00 ± 4.79 | 0.03 | 0.957 |
| <i>MADRS (11:230)</i> | 20.91 ± 12.98 | 18.69 ± 9.16 | 0.650 | 0.421 |
| <i>NMDAR ANTIBODY (IgG; LIVE ASSAY)</i> | | | | |

| | | | | |
|--|------------------|-------------------|-------|-------|
| <i>AVLT IR total</i> (9:158) | 58.00 ± 13.90 | 62.86 ± 12.03 | 0.929 | 0.337 |
| <i>AVLT IR total (CAARMS adjusted)</i> (9:123) | 58.00 ± 13.90 | 62.15 ± 12.13 | 0.826 | 0.365 |
| <i>WAIS estimated total IQ</i> (12:219) | 95.00 ± 19.98 | 98.32 ± 17.10 | 0.067 | 0.796 |
| <i>WAIS estimated total IQ (CAARMS adjusted)</i> (12:177) | 95.00 ± 19.98 | 100.01 ± 17.11 | 0.353 | 0.553 |
| <i>CAARMS total</i> (13:194) | 51.00 ± 11.90 | 49.37 ± 15.21 | 0.197 | 0.657 |
| <i>BPRS positive sx</i> (13:222) | 14.62 ± 4.82 | 13.56 ± 4.57 | 0.683 | 0.409 |
| <i>SANS</i> (13:206) | 15.31 ± 11.39 | 15.76 ± 11.96 | 0.058 | 0.810 |
| <i>YMRS</i> (12:224) | 6.50 ± 5.16 | 3.89 ± 4.76 | 3.179 | 0.076 |
| <i>MADRS</i> (13:228) | 23.69 ± 9.05 | 18.51 ± 9.30 | 3.847 | 0.051 |

Table 3: Cognitive function and psychopathology scores by NMDAR antibody serostatus (descriptive means shown with F and p values from ANOVA). Numbers in brackets under variable names are the number of seropositive subjects and seronegative subjects for whom data were available for that variable. BPRS total score excluded from analysis due to heteroscedasticity.

Abbreviations: NMDAR: N-methyl-d-aspartate receptor, AVLT: Rey Auditory Verbal Learning Task, IR: immediate recall, CAARMS: Comprehensive Assessment of At-Risk Mental States, WAIS: Wechsler Adult Intelligence Scale III, BPRS: Brief Psychiatric Rating Scale, SANS: Scale for the Assessment of Negative Symptoms, YMRS: Young Mania Rating Scale, MADRS: Montgomery-Asberg Depression Rating Scale

Figure 1

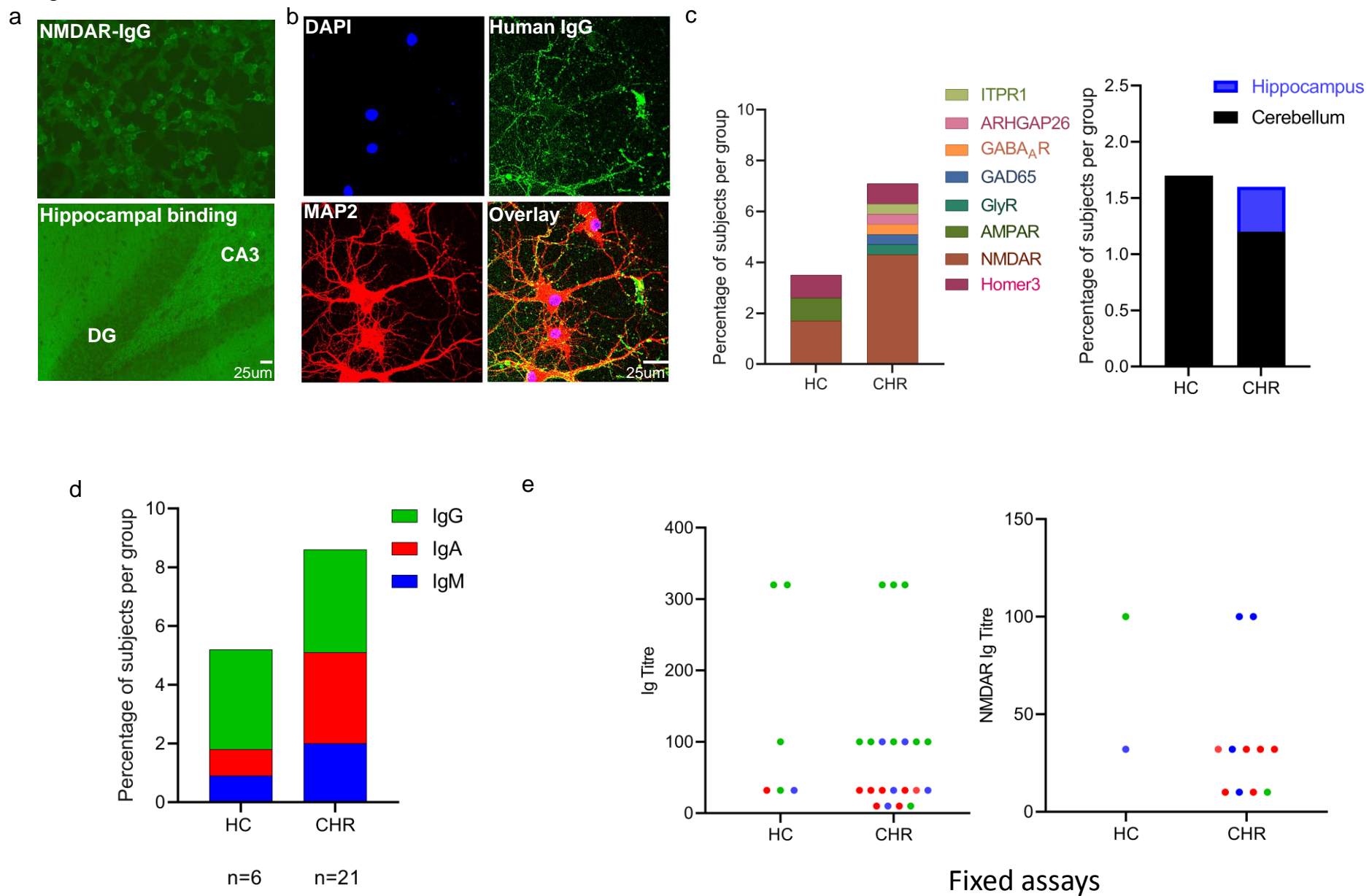


Figure 2

